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# Single-molecule imaging reveals topological isomer-dependent diffusion by 4-armed star and dicyclic 8-shaped polymers†

Satoshi Habuchi,\*a Susumu Fujiwara, b Takuya Yamamoto and Yasuyuki Tezuka\*b

Diffusion dynamics of topological isomers of polymer molecules was investigated at the single-molecular level in the melt state by employing the fluorophore-incorporated 4-armed star and the corresponding doubly-cyclized, 8-shaped poly(THF) chains. While the single-molecule fluorescence imaging experiment revealed that the diffusion of the 4-armed star polymer was described by a single Gaussian distribution, the diffusion of the 8-shaped polymer exhibited a double Gaussian distribution behaviour. We reasoned that the two 8-shaped polymeric isomers have distinct diffusion modes in the melt state, although ensemble-averaged experimental methods cannot detect differences in the overall conformational state of the isomers. The single-molecule experiments suggested that one of the 8-shaped polymeric isomers, having the horizontally oriented form, causes an efficient threading with the linear matrix chains which leads to slower diffusion compared with the corresponding 4-armed star polymer, while the other 8-shaped polymeric isomer, having the vertically oriented form, displayed faster diffusion by the suppression of effective threading with the linear matrix chains due to its contracted chain conformation.

### Introduction

Diffusion and relaxation of polymer chains are the key factors in determining their rheological properties, which serve as a basis for the development of polymer materials such as thin films and fibres. In contrast to linear polymers whose chain dynamics have been well characterized, topological polymers such as ring and dicyclic polymers often display unexpected static and dynamic behaviours. For instance, it was recently demonstrated that the thermal and salt stability of the micelles formed from a polymer increased significantly by a linear-to-cyclic conversion. Such a topology effect should arise from the unique inter- and intra-chain interactions of cyclic polymers which are originated from the absence of chain ends. In addition to the topology dependent chain interactions, topological polymers often adopt different iso-

While ensemble averaged experimental methods have been the primary tools to characterize diffusive and relaxation dynamics of polymers, 12-14 recent single-molecule imaging studies have demonstrated spatiotemporal heterogeneities of polymer dynamics. 15-21 Importantly, those observations cannot be fully interpreted by the current polymer physics theory. In particular, given the unusual and complicated chain dynamics and interactions in topological polymers, single-molecule approaches provide a powerful means for investigating their dynamics. We have recently reported a topology dependent diffusion of individual linear and cyclic polymers in a semidilute solution and in the melt state by employing linear and cyclic poly(THF)s containing a perylene diimide moiety as a fluorophore. 23-25 The single-molecule imaging experiments have revealed an unexpected multiple-mode

meric forms that do not exist in linear and branched polymers because of their unique structural configurations. Such topological isomers often display distinct physical properties such as the isomer-dependent hydrodynamic volume. This could have significant effects on their diffusive and relaxation dynamics, and thus could contribute to the development of new polymer materials through a topological isomer-dependent control of the rheological properties. However, molecular mechanisms of topology- and topological isomer-dependent diffusion and relaxation have not been well understood despite the extensive theoretical and experimental studies.

<sup>&</sup>lt;sup>a</sup>Biological and Environmental Sciences and Engineering, King Abdullah University of Science and Technology, P.O. Box 4700 KAUST, Thuwal 23955-6900, Kingdom of Saudi Arabia. E-mail: satoshi.habuchi@kaust.edu.sa

<sup>&</sup>lt;sup>b</sup>Department of Organic and Polymeric Materials, Tokyo Institute of Technology, 2-12-1 O-okayama, Meguro-ku, Tokyo 152-8552, Japan. E-mail: ytezuka@o.cc.titech.ac.jp † Electronic supplementary information (ESI) available: Calculation of the radius of gyration, cumulative distribution function analysis, and movies of single-molecule diffusion. See DOI: 10.1039/c5py00534e

diffusion of the cyclic polymer in the semidilute solution due to partial threading of the cyclic chains with the linear matrix chains.<sup>23</sup> The study clearly demonstrated the topology effect on the diffusion dynamics of polymers that was often hidden behind the ensemble averaging in the conventional ensemble experiments.

In the present study, we focused on the diffusive dynamics of topological isomers of polymer molecules in the melt state at the single-molecular level as this could be a critical factor for controlling polymer dynamics while only a little is known about their fundamental physical properties. As a model system of topological isomers, we synthesized a pair of 4-armed star (1) and dicyclic 8-shaped polymers.<sup>26</sup> In order to fluorescently visualize the diffusive dynamics of the entire polymers, we have introduced a perylene diimide fluorophore unit commonly in the core junction position of the 4-armed star and dicyclic 8-shaped polymer structures by an electrostatic self-assembly and covalent fixation (ESA-CF) process, which has been reported previously (Fig. 1). 10,27,28 The 8-shaped polymer employed in the present study is considered as an isomeric mixture having vertically oriented (2a) and horizontally oriented (2b) forms (Fig. 1(b)), as reasonably presumed from the previous studies on the double cyclization by

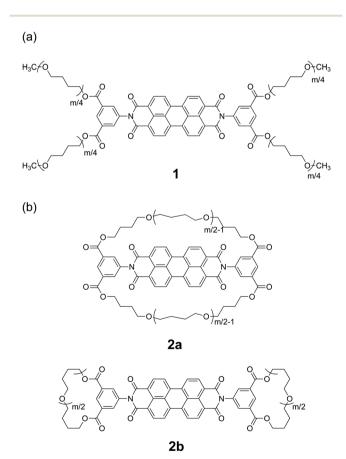


Fig. 1 The 4-armed star (1), vertically oriented 8-shaped dicyclic (2a), and horizontally oriented 8-shaped dicyclic (2b) poly(THF) molecules containing a perylene diimide moiety.

H-shaped polymer precursors.<sup>29</sup> Importantly, the presence of two distinguishable polymeric components could not be substantiated unequivocally through NMR or SEC measurements. Now upon a single-molecule fluorescence imaging study on the set of a 4-armed star polymer and a pair of isomeric dicyclic 8-shaped poly(THF) incorporating a perylene diimide fluorophore, we demonstrate remarkable topology and isomer effects on their dynamics, in particular undetectable through any ensemble methods (*e.g.* NMR, SEC).

# Experimental

#### Sample preparation

1, 2a, and 2b were synthesized using an electrostatic selfassembly and covalent fixation process, which is reported elsewhere. 10,27,28 1 and 2a + 2b have number average molecular weights of  $M_n = 8300 \ (M_w/M_n = 1.04)$  and  $M_n = 12\ 500 \ (M_w/M_n = 1.04)$ 1.10), respectively.<sup>26</sup> Samples for the single-molecule fluorescence imaging experiments were prepared by mixing a melt of unlabelled linear poly(THF) (Aldrich,  $M_n = 3000$ ,  $M_w/M_n =$ 1.89) and a chloroform solution of 1 (1  $\times$  10<sup>-6</sup> M) or 2a + 2b  $(1 \times 10^{-6} \text{ M})$  at a volume ratio of 99 to 1. Then, the solvent was evaporated completely by heating the mixture. The extremely low molar fraction of 1 and 2a + 2b in the melt samples (32-35 ppb) allows spatial isolation of the fluorophore incorporated molecules that is required for the single-molecule analysis of diffusion. A 10 µL volume of the sample was sandwiched between two clean microscope coverslips. The thickness of the melt sample was approximately 10 μm.

#### Single-molecule fluorescence imaging experiment

Single-molecule fluorescence images were recorded by a homebuilt imaging setup which consisted of an inverted microscope (Olympus, IX71) equipped with a high N.A. objective lens (Olympus, ×100, N.A. = 1.3) and EM-CCD camera (Andor Technology, iXon<sup>EM</sup>+).<sup>23</sup> The temperature of the sample was set to 303 K using an objective heater (Bioptechs, temperature stability: ±0.2 °C). 25 The sample was set on the microscope stage and kept at 303 K for 1 hour before recording the fluorescence images. This ensures that the sample has reached the equilibrium conditions. Circularly-polarized light from a 488 nm line of a CW Ar-Kr ion laser (Coherent Innova 70C) obtained by a Berek compensator (Newport) was used for excitation. The excitation light was introduced into the microscope through an excitation filter (Semrock, LL01-488-12.5), reflected by a dichroic mirror (Omega optical, 500DRLP), and the sample was illuminated through the objective. We adjusted an illumination area to approximately 20 µm in diameter. The average excitation power was 2 kW cm<sup>-2</sup>. The fluorescence from the sample was corrected by the same objective, passed through the dichroic mirror and an emission filter (Semrock, BLP01-488R-25), and was detected by the EM-CCD camera. The fluorescence images were recorded at 133 Hz (i.e., 7.5 ms exposure time). The pixel size of the images was 160 nm.

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#### Image analysis

The diffusion rate and mode of 1 and 2a + 2b were analysed by using a single-molecule localization and tracking method. First, the locations of the molecules in each image frame were precisely determined by two-dimensional Gaussian fitting using routines written in MATLAB. One were determined by mean-squared displacement (MSD) analysis of the trajectories. In order to compare experimentally determined D distributions with theoretically calculated probability distributions, the diffusion trajectories were all cut such that they contained ten data points (see below).

#### Cumulative distribution function (CDF) analysis

The modes of diffusion were analysed using a CDF,  $P(r^2, i\Delta t)$ , which corresponds to the cumulative probability of finding the diffusing molecule within a radius r from the origin after a certain time lag  $i\Delta t$ .

**Homogeneous diffusion model.** CDFs obtained for molecules that show homogeneous diffusion with a distinct *D* value were fitted by the equation, <sup>32</sup>

$$P(r^2, i\Delta t) = \int_0^r (r^{2'}, i\Delta t) dr' = 1 - A \exp\left[-\frac{r^2}{4D(i\Delta t)}\right]$$
(1)

any deviation from a single-exponential decaying behaviour suggests the heterogeneous diffusion of the molecules in the sample. The CDF coefficients  $(r^2)$  at different time lags were determined by fitting the CDFs with eqn (1). The D value was estimated by the slope of the  $r^2$  plot.

Gaussian distribution model. CDFs obtained for molecules that show multiple diffusion modes in which the D value distribution was described by a Gaussian were fitted by the equation,  $^{25}$ 

$$1 - P(r^2, i\Delta t) = \int_0^\infty f(D)D^{-1} \exp\left[-\frac{r^2}{4D(i\Delta t)}\right] dD \qquad (2)$$

$$f(D) = A \exp\left\{-\frac{(D - D_0)^2}{2w^2}\right\}$$
 (3)

where f(D) denotes probability distribution of D described by a Gaussian. A, w, and  $D_0$  are the amplitude, width, and centre of the Gaussian, and those values were determined by the fitting. The  $r^2$  values at different time lags were determined by fitting the CDFs with eqn (2). The mean D value was estimated by the slope of the  $r^2$  plot.

**Double Gaussian model.** CDFs obtained for molecules that show multiple diffusion modes in which the *D* value distribution was described by multiple continuous and broad distribution (*i.e.*, multiple Gaussian) were fitted by the equation,

$$1 - P(r^2, i\Delta t) = \sum A_j \int_0^\infty f(D_j) D_j^{-1} \exp\left[-\frac{r^2}{4D_j(i\Delta t)}\right] dD_j \quad (4)$$

$$f(D_j) = \alpha_j \exp\left\{-\frac{\left(D_j - D_{0j}\right)^2}{2w_j^2}\right\}$$
 (5)

where  $f(D_j)$  denotes probability distribution of the  $j^{\text{th}}$  component of D described by a Gaussian.  $A_j$  is the fraction of each diffusion component.  $\alpha_j$ ,  $w_j$ , and  $D_{0j}$  are the amplitude, width, and centre of the  $j^{\text{th}}$  component of the Gaussian. The  $r^2$  values of different diffusion modes at different time lags were determined by fitting the CDFs with eqn (4). The mean D values for each diffusion mode were estimated by the slope of the  $r^2$  plot.

#### Statistical probability distribution of the diffusion coefficient

**Homogeneous diffusion model.** The statistical probability distribution of the diffusion coefficient in a homogeneous environment was calculated using the following equation,<sup>33</sup>

$$p(D)dD = \frac{1}{(N-1)!} \left(\frac{N}{D_0}\right)^N D^{N-1} \exp\left(\frac{-ND}{D_0}\right) dD$$
 (6)

where N is the number of the data points in a diffusion trajectory,  $D_0$  is the mean diffusion coefficient, and D is the experimentally obtained diffusion coefficient for an individual trajectory. For this calculation, the experimentally obtained trajectories were all cut such that they contained ten data points (*i.e.*, N = 10). The  $D_0$  value for the calculation was determined by the CDF analysis of the diffusion data (the D value in eqn (1)).

**Gaussian distribution model.** When the diffusion is described by multiple modes in which the *D* value distribution is described by a Gaussian, the statistical probability distribution was calculated using the following equation, <sup>25</sup>

$$p(D) = \int_{0}^{\infty} f(D) \frac{1}{(N-1)!} \left(\frac{N}{D_0}\right)^{N} D^{N-1} \exp\left(\frac{-ND}{D_0}\right) dD \qquad (7)$$

where f(D) denotes probability distribution of D determined by the CDF analysis of the diffusion data using eqn (2) and (3).  $D_0$  was also determined by fitting the CDFs with eqn (2) and (3).

**Double Gaussian model.** When the diffusion is described by multiple diffusion modes in which the *D* value distribution is described by multiple Gaussian, the statistical probability distribution was calculated using the following equation,

$$p(D) = \sum_{j=0}^{\infty} f(D_j) \frac{1}{(N-1)!} \left(\frac{N}{D_{0j}}\right)^N D_j^{N-1} \exp\left(\frac{-ND_j}{D_{0j}}\right) dD_j$$
 (8)

where  $f(D_j)$  denotes probability distribution of the  $f^{th}$  component of  $D(D_j)$  determined by the CDF analysis of the diffusion data using eqn (4) and (5).  $D_{0j}$  is the mean diffusion coefficient of the  $f^{th}$  component determined by fitting the CDFs with eqn (4) and (5).

#### Results and discussion

Absorption and fluorescence spectra of **1** and **2a** + **2b** in THF are virtually identical (Fig. 2), with absorption and fluorescence maxima at 523 and 532 nm, respectively. Fluorescence

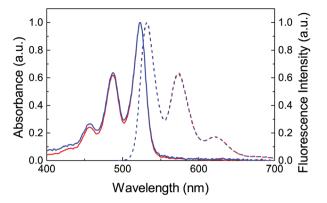


Fig. 2 Absorption (solid lines) and fluorescence (broken line) spectra of 1 (red lines) and 2a + 2b (blue lines) in THF. The excitation wavelength of the fluorescence spectra was 488 nm.

rescence quantum yields of 1 (0.46 in THF) and 2a + 2b (0.52 in THF) are comparable to that of a precursor perylene diimide derivative (0.54 in ethanol).<sup>23</sup> The results confirm that the poly(THF) chains have a minimum effect on the fluorescence properties of the fluorophore.

Fig. 3 shows single-molecule fluorescence images of 1 (Fig. 3(a)) and 2a + 2b (Fig. 3(b)) in the melt of the linear poly-(THF) matrix recorded at 303 K. A bright fluorescence of a single perylene diimide fluorophore incorporated into the chain was observed which confirmed that the centre-of-mass diffusion of individual polymer molecules in the melt was directly visualized. The spatial locations of the molecule in each image were determined by fitting the image to a twodimensional Gaussian function (see the Experimental section). Examples of the diffusion trajectories obtained by the image analysis are shown in Fig. 3(a) and (b) in yellow. The diffusion coefficients of each molecule determined by the MSD analysis of the diffusion trajectories<sup>23</sup> are plotted in frequency histograms (Fig. 4 grey bars). The D distributions obtained in Fig. 4 are much broader than those calculated theoretically for a homogeneous diffusion of the polymer chains due to the statistical error of the analysis of the diffusion trajectories (Fig. 4 green lines, see the Experimental section: Statistical prob-

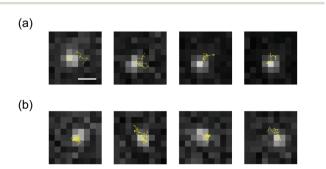


Fig. 3 Single-molecule fluorescence images of (a) 1 and (b) 2a+2b each mixed with linear poly(THF) melt. The yellow lines show diffusion trajectories. Scale bar =  $0.5~\mu m$ .

ability distribution of the diffusion coefficient – homogeneous diffusion model), suggesting heterogeneous diffusion of 1 and 2a + 2b in the melt state. Similar heterogeneous diffusion has been observed for a linear and mono-cyclic poly(THF) in the melt state in the previous study.<sup>25</sup>

The diffusion mode of 1 and 2a + 2b was further analysed by the cumulative distribution functions (CDFs), P  $(r^2, i\Delta t)$ .<sup>32</sup> A single-exponential and multi-exponential decaying behaviour of CDFs in the form of 1-P corresponds to a single-mode and multi-mode diffusion, respectively (see the Experimental section: Cumulative distribution function analysis). The CDFs obtained for 1 and 2a + 2b show multi-exponential decaying behaviour (Fig. 5(a) and (b)). The results demonstrate clearly the heterogeneous diffusion of 1 and 2a + 2b in the melt state. The diffusion of the linear and mono-cyclic poly(THF)s was well-described by a broad and continuous distribution of the diffusion coefficient of each polymer chain due to the different numbers of entanglement in each polymer chain.<sup>25</sup> The CDFs obtained for 1 were fitted well by this model (Fig. 5(a), see the Experimental section: Cumulative distribution function analysis - Gaussian distribution model). The frequency histogram of the D values obtained from 1 was also reasonably reproduced by the model (Fig. 4(a) red line, see the Experimental section: Statistical probability distribution of diffusion coefficient - Gaussian diffusion model). Furthermore, the D value determined by the CDF coefficients ( $D = 0.125 \mu \text{m}^2 \text{ s}^{-1}$ , Fig. 5(c)) is in good agreement with the median of the D distribution obtained in the MSD analysis ( $D = 0.133 \mu \text{m}^2 \text{ s}^{-1}$ , Fig. 4(a)). All these results strongly suggest that the 4-armed star polymer (1) shows broad and continuous distribution of the diffusion coefficient in the melt state that is described by a Gaussian function.

The mean diffusion coefficient of the 4-armed star polymer (1) in the melt state ( $D=0.133~\mu\text{m}^2~\text{s}^{-1}$ ) is slower than that of the linear (*i.e.*, 2-armed) polymer ( $D=0.224~\mu\text{m}^2~\text{s}^{-1}$ ) with a similar arm length reported previously. Although theoretical description of the diffusion behaviour of those polymers based on the reptation model is not straightforward due to the relatively small number of the chain interaction in each arm, the difference in the mean D values can be interpreted by the armnumber dependent diffusion of branched polymers.<sup>5</sup>

In contrast to the 4-armed star polymer (1), the diffusion mode of the isomeric dicyclic 8-shaped polymer (2a + 2b) cannot be described by the single Gaussian distribution model. Although the CDFs were fitted well by the single Gaussian model (see Fig. S1†), the frequency histogram of the D values showed systematic deviation from the D distribution calculated theoretically based on the CDF analysis (Fig. 4(b) red line, see the Experimental section: Statistical probability distribution of diffusion coefficient – Gaussian diffusion model). Moreover, the D value determined by the CDF coefficients (D = 0.089  $\mu$ m<sup>2</sup> s<sup>-1</sup>, Fig. S1†) is not in good agreement with the median of the D distribution obtained in the MSD analysis (D = 0.156  $\mu$ m<sup>2</sup> s<sup>-1</sup>, Fig. 4(b)). These results suggest that the single Gaussian distribution model does not properly describe the diffusion of 2a + 2b in the melt state.

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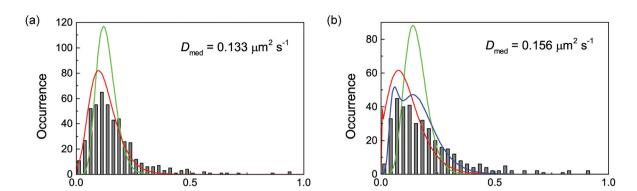


Fig. 4 Frequency histograms (grey bars) of the diffusion coefficient determined for (a) 1 and for (b) 2a + 2b each mixed with linear poly(THF) melt. The solid lines show theoretically calculated probability distributions of the diffusion coefficient based on three different diffusion models; homogeneous diffusion model (green lines, eqn (6)), Gaussian distribution of the diffusion coefficient (single Gaussian model, red lines, eqn (7)), and two distinct diffusion modes whose distributions are described by Gaussian functions (double Gaussian model, blue line, eqn (8)). The distributions of the diffusion coefficients and the relative amplitudes used in the calculations were determined by the fitting of the cumulative distribution functions with eqn (1) (homogeneous diffusion model), eqn (2) (single Gaussian model), and eqn (4) (double Gaussian model).

 $D (\mu m^2 s^{-1})$ 

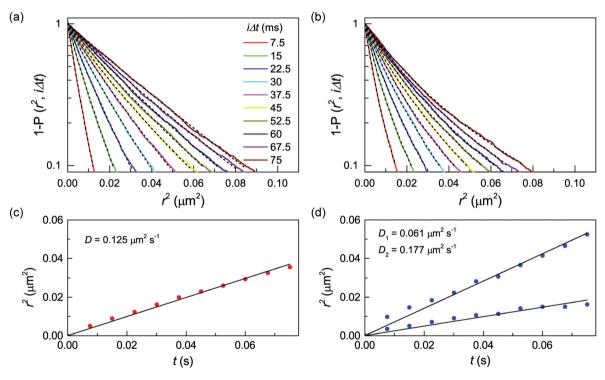


Fig. 5 Experimentally obtained cumulative distribution functions ( $i\Delta t = 7.5-75$  ms) in the form of 1-P (solid lines) for (a) 1 and for (b) 2a + 2b. Dashed lines show fittings with (a) eqn (2) and (b) eqn (4). CDF coefficients at different time lags for (c) 1 and for (d) 2a + 2b. The *D* values for each diffusion mode were determined by either single Gaussian distribution (red) or double Gaussian distribution (blue) models using eqn (2) and (4), respectively. The solid lines show linear fittings.

Two isomeric forms, vertically oriented (2a) and horizontally oriented (2b) 8-shaped dicyclic polymers, could result in two distinct diffusion modes. Thus, the CDFs obtained from 2a + 2b were fitted by a double Gaussian distribution model in which the diffusion is described by two independent broad and continuous distributions (see the Experimental section:

Cumulative distribution function analysis – Double Gaussian model). The CDFs can be fitted well with this model (Fig. 5(b)). The frequency histogram of the D values is also reasonably reproduced by the theoretically calculated D distribution based on the model (Fig. 4(b) blue line, see the Experimental section: Calculation of the statistical probability distribution

 $D (\mu m^2 s^{-1})$ 

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of diffusion coefficient – Double Gaussian model). Those results strongly suggest the presence of two distinct diffusion modes while we cannot rule out completely the possibility of having more than two modes. Two diffusion coefficients ( $D = 0.061 \ \mu\text{m}^2 \ \text{s}^{-1}$  and  $D = 0.177 \ \mu\text{m}^2 \ \text{s}^{-1}$ ) were obtained by the linear fits of the CDF coefficients (Fig. 5(d) blue circles). The ratio of the fast to slow diffusing components is estimated to be four to one based on the double Gaussian analysis (Fig. 4(b)).

Although NMR and SEC measurements on 2a + 2b showed no sign of conformational differences between the two isomers,  $^{26}$  the single-molecule fluorescence imaging experiment suggests distinct diffusion dynamics of the two topological isomers. As the dimension of the perylene diimide fluorophore unit (1.1 nm length) is nearly a half of the radius of gyration ( $R_{\rm G}$ ) of the poly(THF) chain in each ring ( $R_{\rm G}$  = 2.4 nm, see the ESI:† calculation of gyration radius), the chain conformation and the interaction with the linear matrix chains should depend on how the dicyclic chains are connected to the fluorophore while the dicyclic chains of poly(THF) in 2a + 2b adopt the overall random coil conformation in both the isomers.

The mean D value of the isomeric dicyclic 8-shaped polymer (2a + 2b) in the melt state ( $D = 0.156 \mu \text{m}^2 \text{ s}^{-1}$ ) is comparable to that of the monocyclic polymer ( $D = 0.155 \mu m^2 s^{-1}$ ) with a similar ring size reported previously, which is in sharp contrast to the 4-armed star polymer (1) whose average diffusion coefficient is interpreted as the arm-number dependent diffusion of branched polymers.<sup>25</sup> Although the number average molecular weight of 2a + 2b determined by NMR ( $M_n =$ 12 500) is much larger than that of the monocyclic polymer  $(M_n = 3800)$ , the peak molecular weight of 2a + 2b determined by SEC  $(M_p = 5600)$  is only slightly larger than that of the monocyclic polymer ( $M_p = 4000$ ). 2a and 2b are structurally similar to  $\theta$ -shaped and manacle-shaped isomers of a dicyclic polymer. Previous studies demonstrated that the hydrodynamic volume of the  $\theta$ -shaped isomer is much smaller than that of the manacle-shaped isomer. 10,11 If the ESA-CF reaction between the poly(THF) and the fluorophore occurs at random probability, the molar ratio between 2a and 2b should be two to one. This suggests that the major component of the dicyclic 8-shaped polymer (i.e., 2a) should have significantly compact conformation as compared with the monocyclic polymer. The results obtained from the SEC experiments are, therefore, consistent with the predicted contracted conformation of the dicyclic polymer,26 which partly explains the small difference in the mean D values between the monocyclic and dicyclic polymers.

The D value of the slower diffusing component observed in  $2\mathbf{a}+2\mathbf{b}$  ( $D=0.061~\mu\text{m}^2~\text{s}^{-1}$ ) is much smaller than that in  $\mathbf{1}$  ( $D=0.133~\mu\text{m}^2~\text{s}^{-1}$ ) although the total chain length in  $2\mathbf{a}+2\mathbf{b}$  is similar to that in  $\mathbf{1}$  (see the ESI:† calculation of gyration radius). A similar trend was observed for the monocyclic and linear (*i.e.*, 2-armed) polymers. <sup>25</sup> The results also coincide with a simulation study which predicts slower diffusion of cyclic chains by the threading with linear chains. <sup>34</sup> The results indi-

cate efficient threading of the dicyclic chains with the linear matrix chains in one isomer of 2a + 2b which results in the slower diffusion of 2a + 2b as compared with 1. Since the slow diffusing isomer in 2a + 2b is a minor component (Fig. 4(b)), this component could be assigned to 2b. In contrast to the slower diffusion component, the D value of the faster diffusing component observed in 2a + 2b ( $D = 0.17 \mu m^2 s^{-1}$ ) is slightly faster than that in 1. Since the total chain length in 1 and in 2a + 2b is comparable, the faster diffusion observed for one of the isomers in 2a + 2b should reflect less efficient threading of 2a + 2b and/or a contracted chain conformation in 2a + 2b as compared with the branched counterpart (1), which is suggested by the SEC measurement. Since 2a is expected to have a contracted chain conformation and represents the major component, this isomer could be responsible for the fast diffusing component. These results suggest that the chain interaction in the melt state is affected not only by the topological state of the chain but also by the isomeric conformation of the chain.

# Conclusion

In this study, we investigated the diffusion modes of the topological isomers in the polymer melt at the single molecular level. The synthesis of the perylene diimide incorporated dicyclic 8-shaped poly(THF) (2a + 2b) provided the unique opportunity to analyse the topological-isomer directed diffusion dynamics. We demonstrated for the first time the distinct diffusion modes of the topological isomers in the melt of the linear matrix polymer by the direct visualization of the diffusive motion of the isomeric dicyclic 8-shaped polymer. Comparisons with the corresponding branched polymer suggested that the two topological isomers were responsible for the two diffusion modes of the dicyclic chain due to their distinct conformational states that lead to isomer-dependent threading with the matrix chains. Such hidden complexity cannot be revealed by ensemble-averaged techniques. This study clearly demonstrated the importance of visualizing individual chains for the molecular level understanding of the topology and isomer dependent polymer dynamics.

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